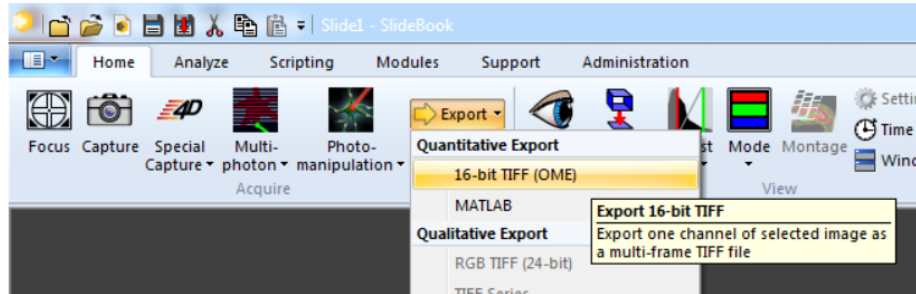


## Steps for Deconvolution processing of diSPIM data with Huygens (SPIM, Deconvolution & Fusion) software.

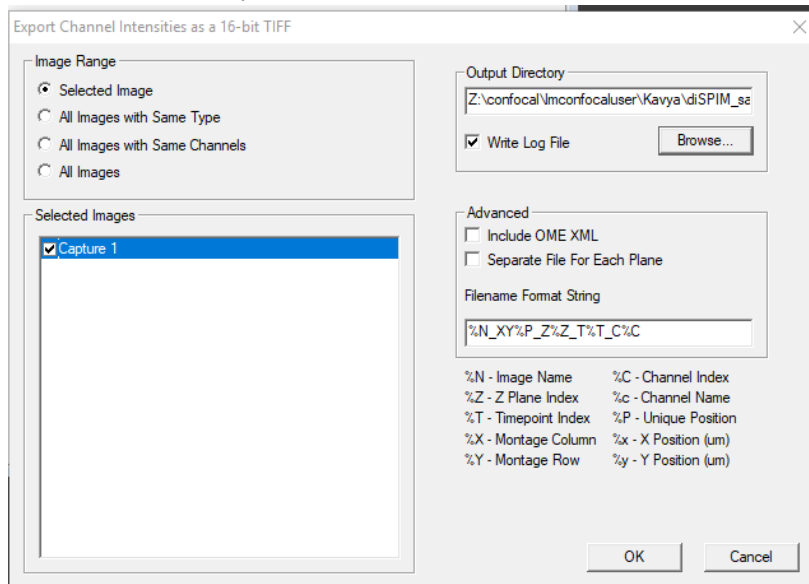
The following steps are applicable for islice data (slice scan – single channel acquisition)

### 1) Export of raw Data from Slidebook:

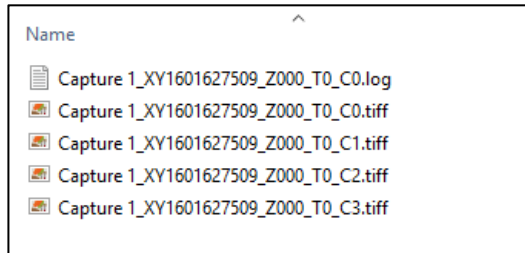
i. Go to Home> Export>16-bit TIFF.



ii. Select the default options and choose the folder (kraken\confocal\lmconfocaluser\...) you would want to export to.



- iii. The exported files consist of .tiff files for each channel, and a .txt file with exported data log.

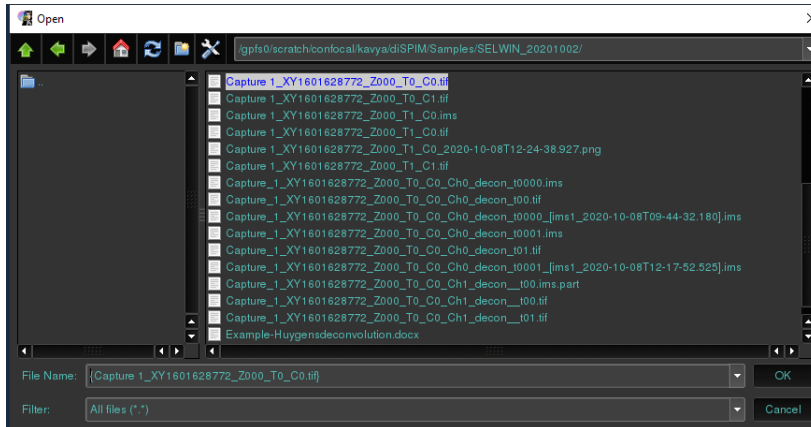


## 2. Processing Deconvolution with Huygens Software:

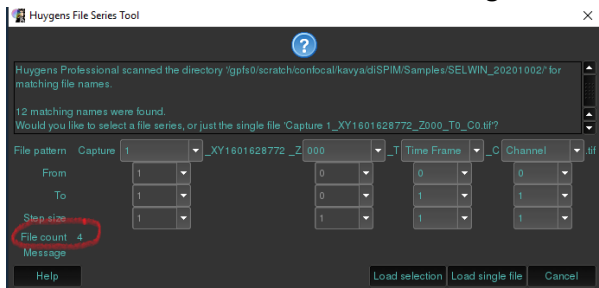
### a) Importing files

- i. File -> Open

Select the first channel Tiff file exported (**C0.tif**).

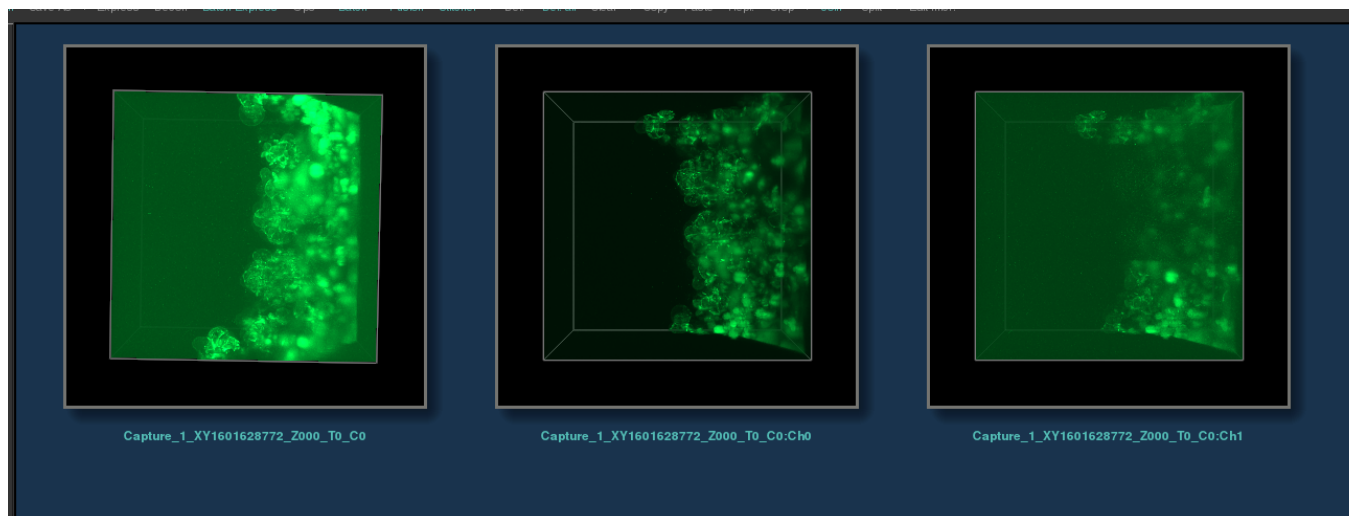
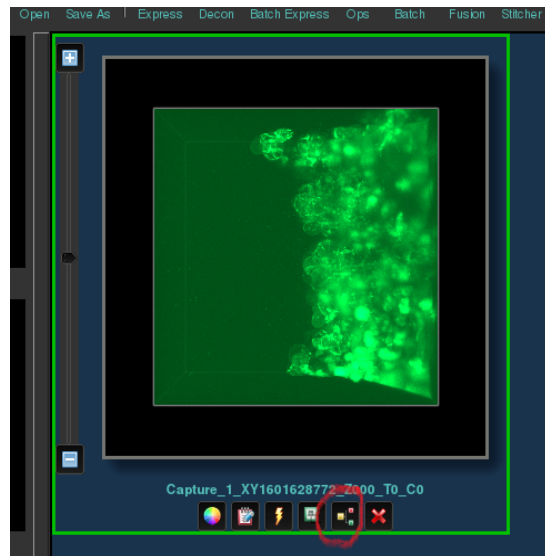


The first channel file C0.tif is a **stack image** that is the image sequence of all the images under the capture file exported.



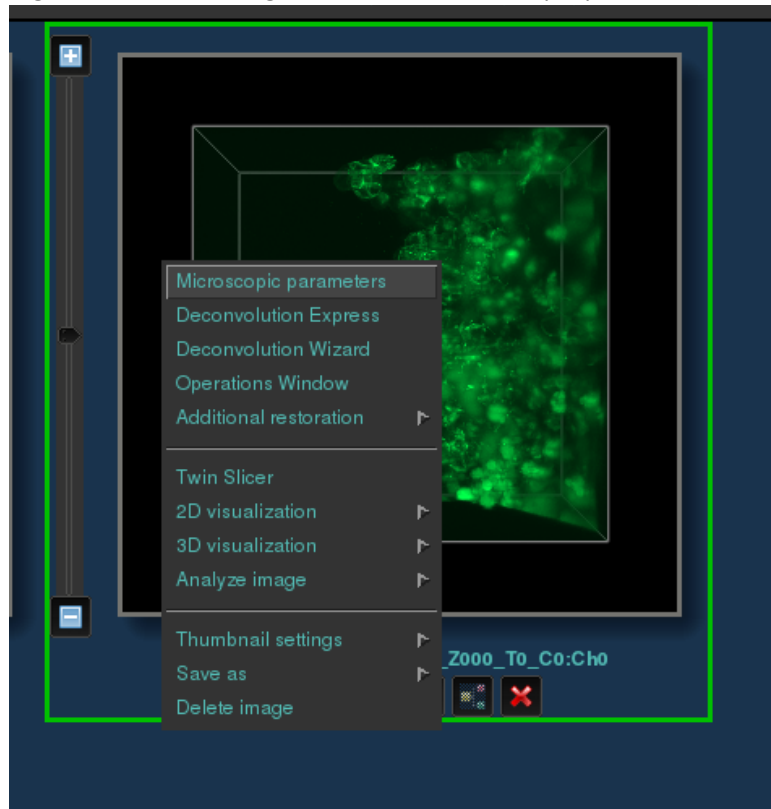
Click 'Load selection' to load all the files in the stack

- ii. After the image stack is opened, click the 'Spilt channels' icon for splitting of the images (channels) for processing

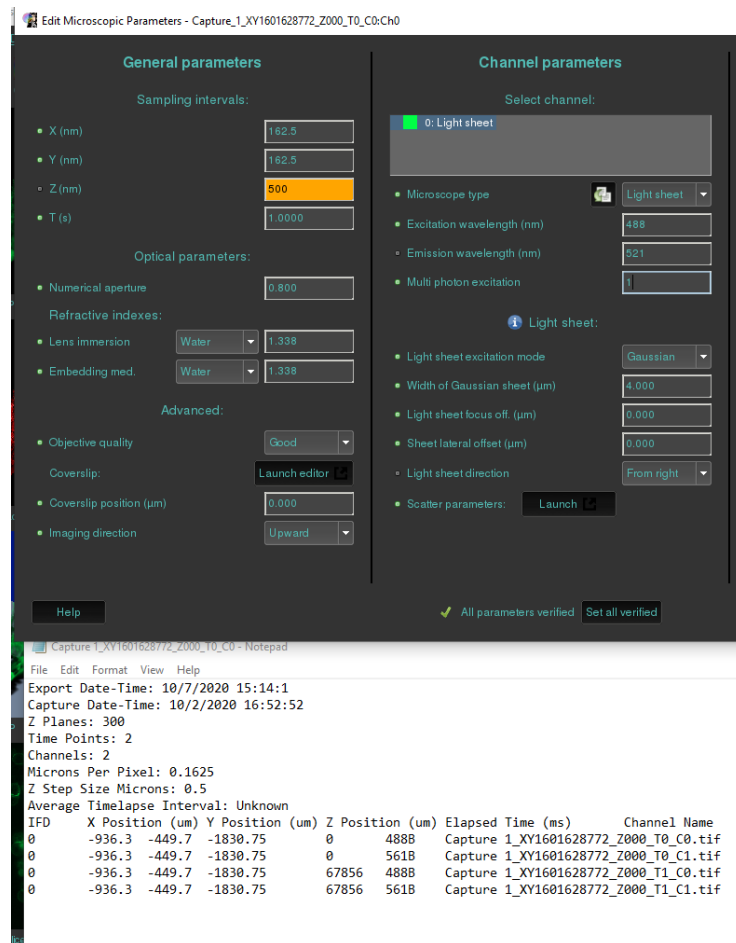


b) Set microscope parameters for the image according to the acquisition protocol.

i. Right click on the image and select 'Microscopic parameters'



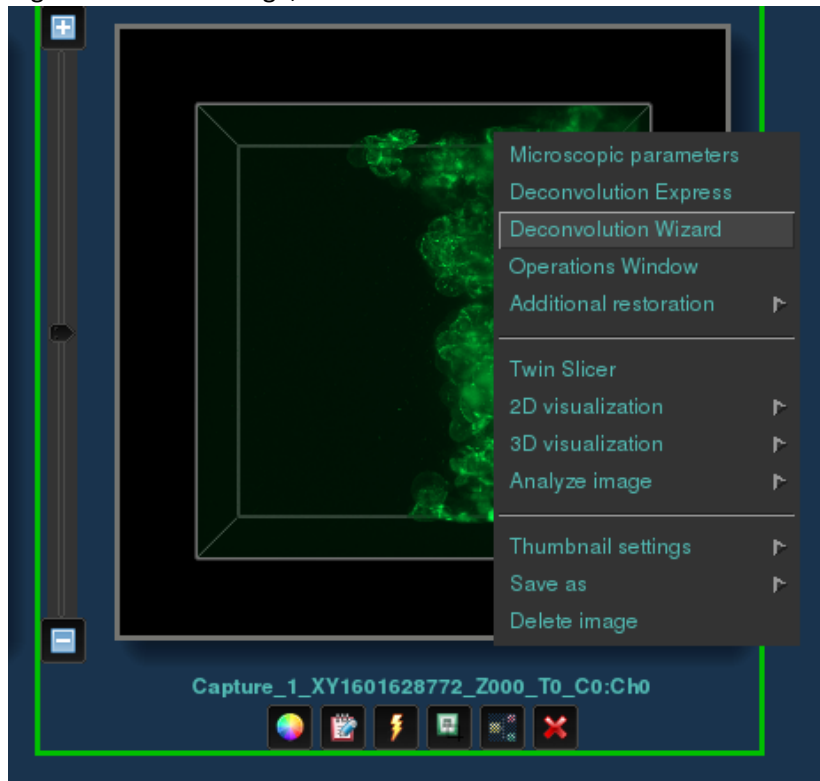
- ii. Enter the parameters based on the acquisition setup. Information for the particular channel can be found in the .txt file (X, Y, Z sampling intervals and wavelength used).
- For diSPIM, X and Y sampling interval = 162.5nm  
 Z sampling interval as set in experiment (found in .txt file). When acquisition ensure z sampling interval <480nm to match Nyquist criterion of sampling  
 For excitation wavelength: 405nm emission wavelength to be set: 440nm  
 For excitation wavelength: 488nm emission wavelength to be set: 521nm  
 For excitation wavelength: 561nm emission wavelength to be set: 601nm  
 For excitation wavelength: 640 nm emission wavelength to be set: 700nm  
 Scattering parameters can be set to 'No scattering' for thin samples without much scattering effect of light.



CHANNEL A: LIGHT SHEET DIRECTION – LEFT  
 CHANNEL B: LIGHT SHEET DIRECTION - RIGHT

c) Using deconvolution wizard

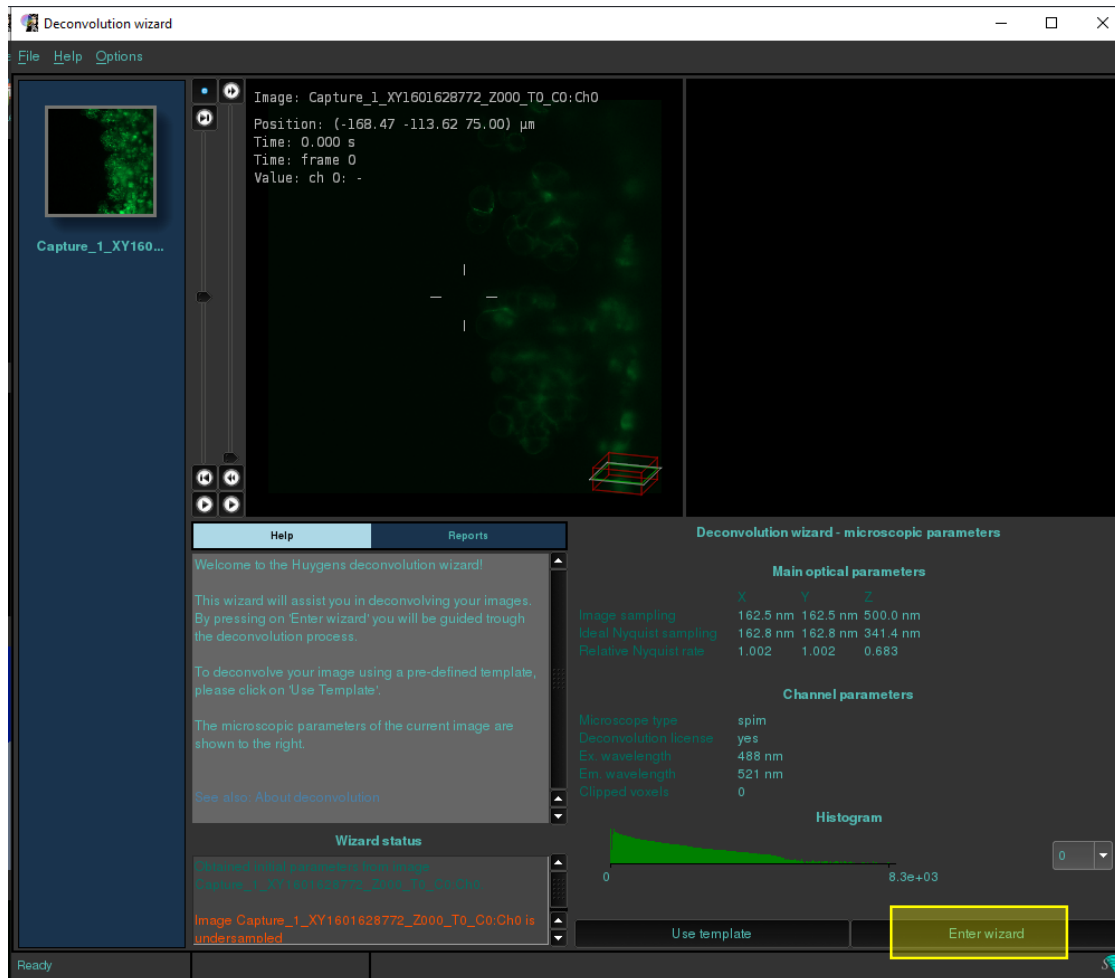
- i. Right click on the image, select 'Deconvolution wizard'



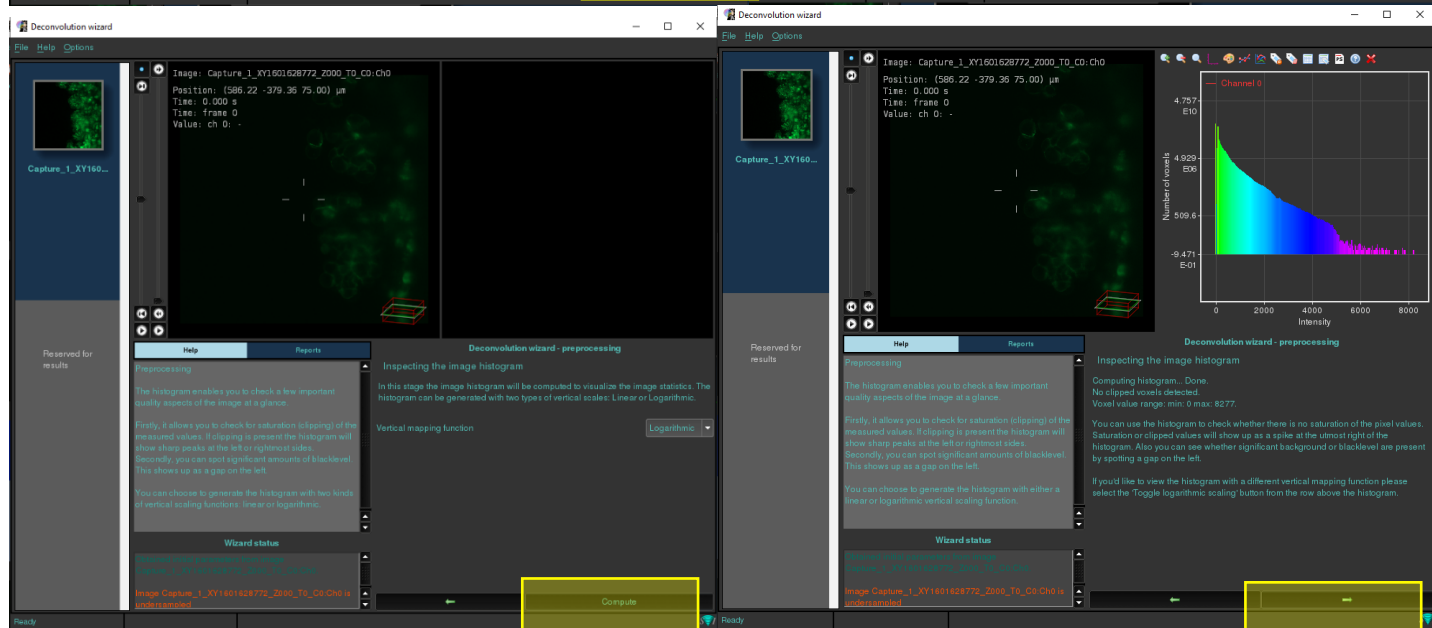
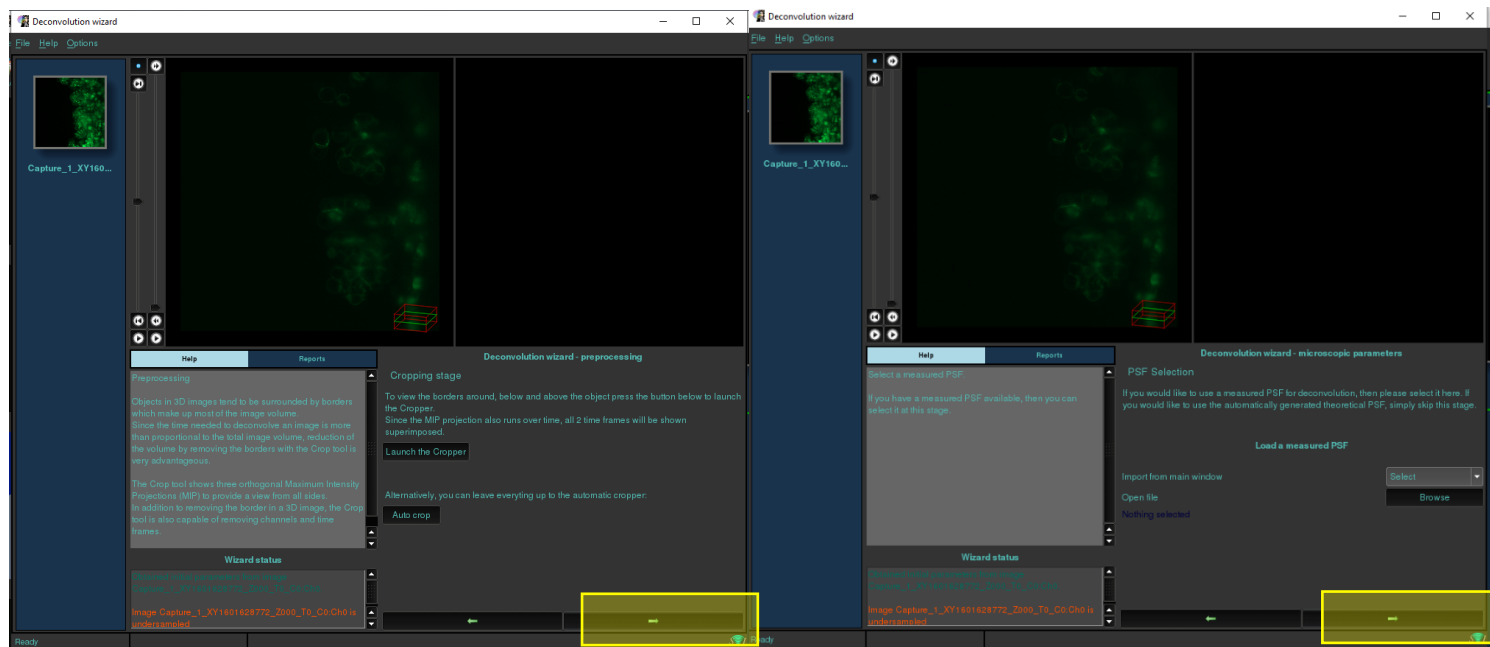
\*Note: For stage scans additional preprocessing are required before deconvolution

- ii. Go through the deconvolution wizard steps. Theoretical PSF is used by default for the deconvolution. Check the microscopic parameters set for the image (X, Y, Z sampling, wavelength information etc.)

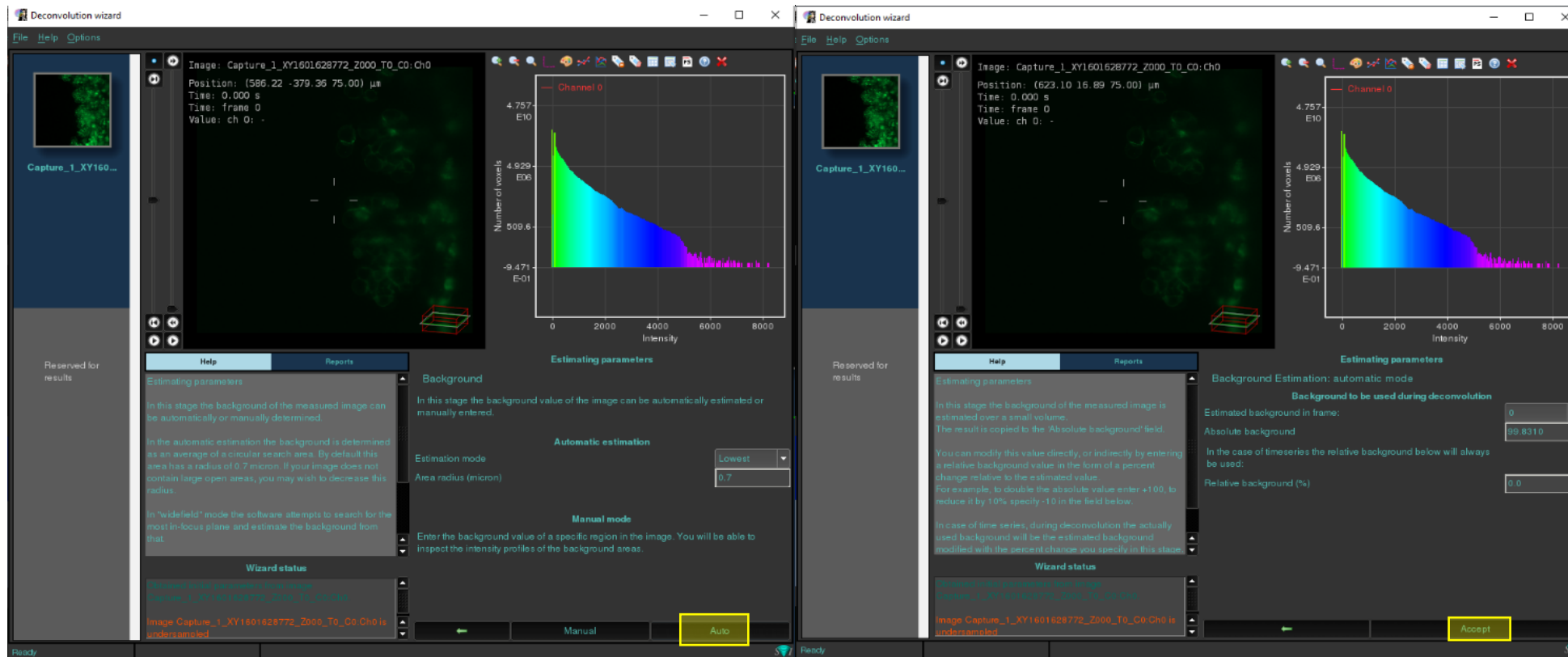
### Enter wizard

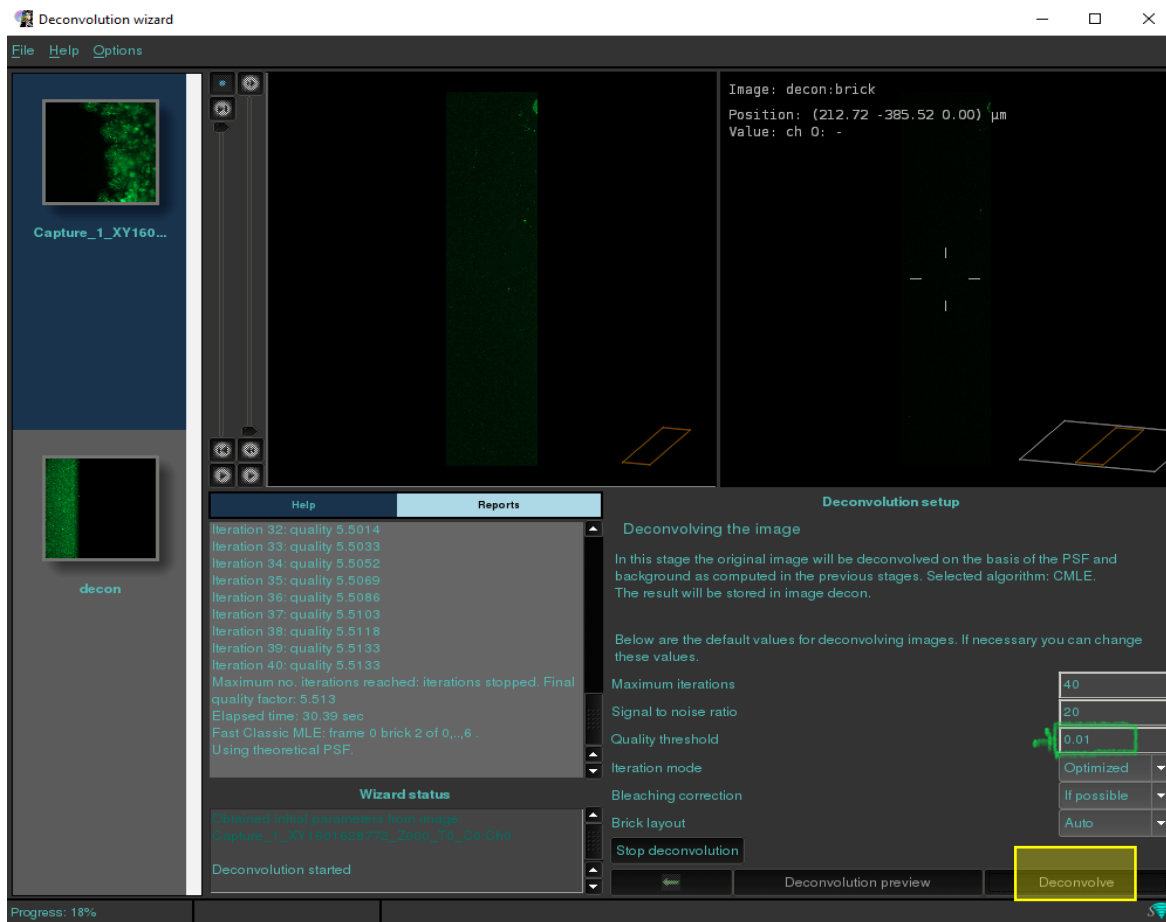


Follow the default steps highlighted in yellow.





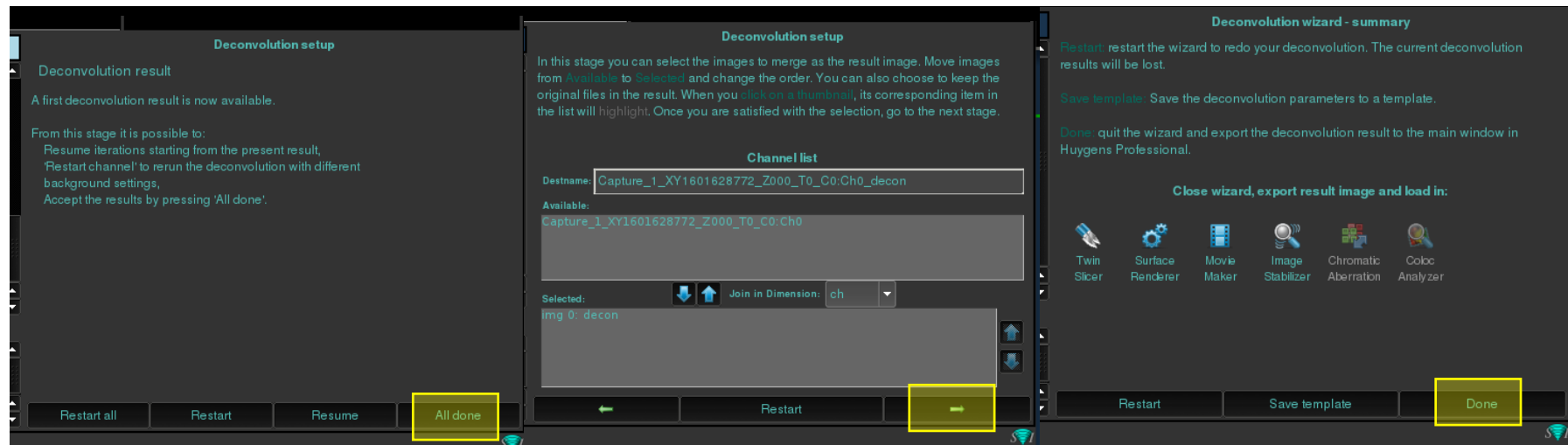




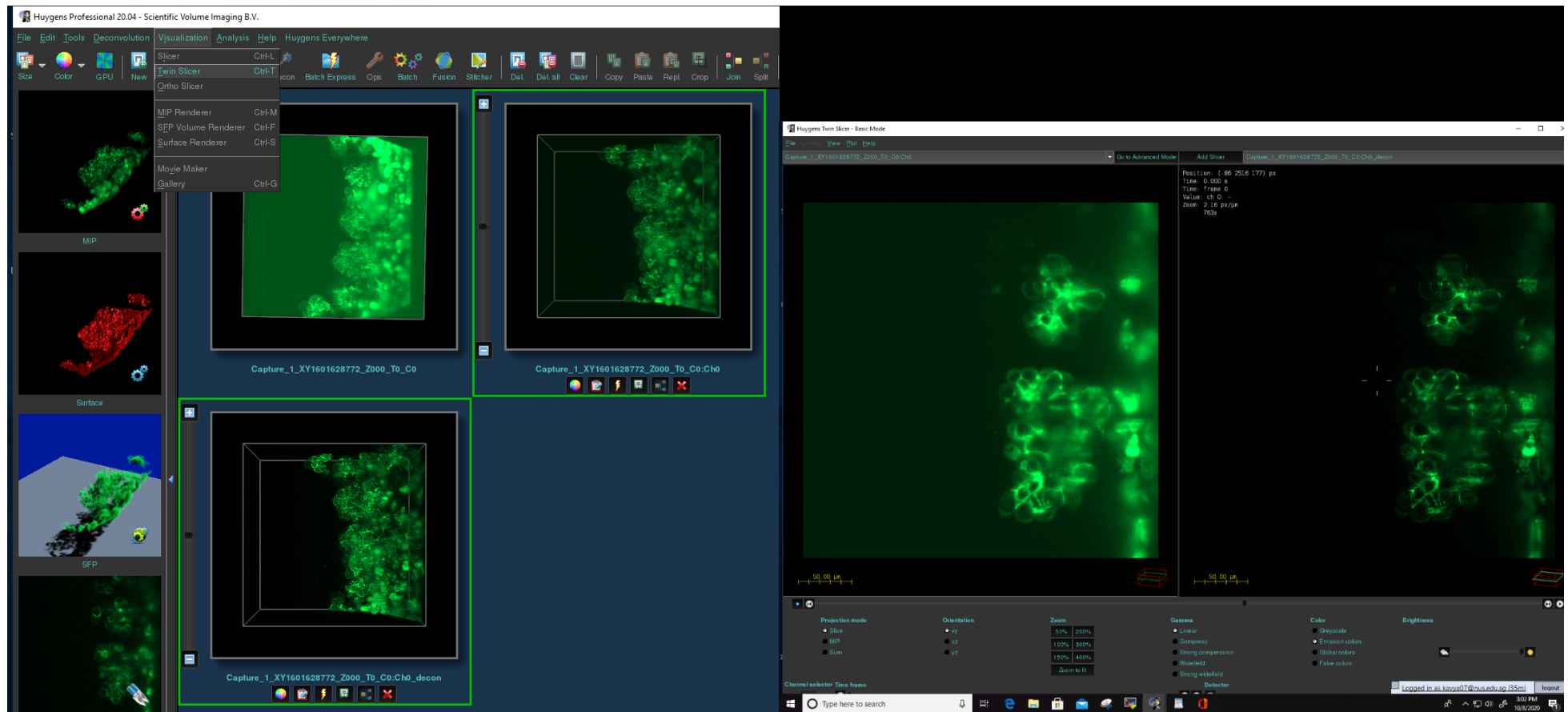
Decrease the quality threshold to 0.01 for better quality of deconvolution.

Other default values are suitable for the sample data.

For further improvisation, increase maximum iteration number and/or change iteration mode to 'Classic'



Name the deconvolved file.



To compare the two images before and after deconvolution, select the two images.

Visualization -> Twin slicer

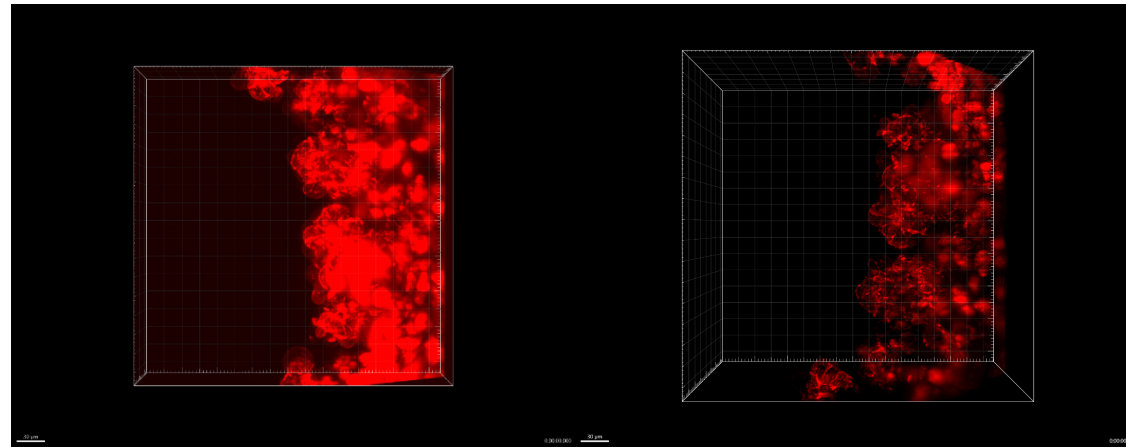
Comparison after deconvolution can be visualized with win slicer.

If deconvolution result is satisfactory, save the file.

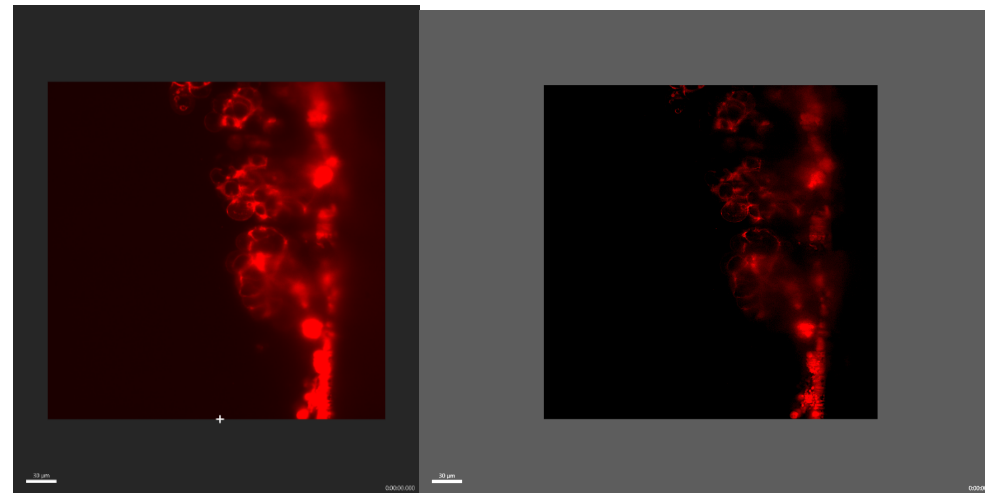
Example image

Deconvolved

3-D volume view



Slice view



Note: The above steps are suitable for slice scans image processing with dispm.

Additional steps

For dislice (slice scans with two orthogonal views) fusion after deconvolution will need to be performed

For istage/ distage scans deskewing of data will need to be performed additionally as pre-processing.